

ISSN 1000-0933
CN 11-2031/Q

生态学报

Acta Ecologica Sinica



第31卷 第15期 Vol.31 No.15 2011

中国生态学学会
中国科学院生态环境研究中心
科学出版社

主办
出版



中国科学院科学出版基金资助出版

生态学报 (SHENTAI XUEBAO)

第31卷 第15期 2011年8月 (半月刊)

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期刊基本参数:CN 11-2031/Q * 1981 * m * 16 * 320 * zh * P * ¥ 70.00 * 1510 * 34 * 2011-08



封面图说:塞罕坝地处内蒙古高原南缘向华北平原的过渡带,地势分为坝上、坝下两部分。解放初期,这里是“飞鸟无栖树,黄沙遮天日”的荒原沙丘,自1962年建立了机械化林场之后,塞罕坝人建起了110多万亩人工林,造就了中国最大的人工林林场。这是让人叹为观止的落叶松人工林海。

彩图提供:陈建伟教授 国家林业局 E-mail: cites.chenjw@163.com

毛轶清, 郑青松, 陈健妙, 刘兆普, 刘国红, 姜超强. 喷施多效唑提高麻疯树幼苗耐盐性的生理机制. 生态学报, 2011, 31(15): 4334-4341.
Mao Y Q, Zheng Q S, Chen J M, Liu Z P, Liu G H, Jiang C Q. Physiological mechanism of foliage spraying paclobutrazol on increasing salt tolerance of *Jatropha curcas* Seedlings. Acta Ecologica Sinica, 2011, 31(15): 4334-4341.

喷施多效唑提高麻疯树幼苗耐盐性的生理机制

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摘要: 研究了 600 mg/L PP₃₃₃ 喷施对 200 mmol/L NaCl 胁迫处理下麻疯树幼苗干重、含水量、叶片细胞超微结构、光合作用、叶片渗透调节能力、叶片丙二醛含量和叶片抗氧化能力的影响。结果表明: 600 mg/L PP₃₃₃ 喷施处理能显著提高 200 mmol/L NaCl 胁迫下植株的干重、根冠比和叶片含水量, 同时显著降低叶片电解质外渗率, 降低叶片细胞超微结构的伤害程度, 显著提高了其叶绿素含量、净光合速率、渗透调节能力、SOD 酶活性和 CAT 酶活性, 显著降低了 MDA 含量和 POD 酶活性。可见, PP₃₃₃ 喷施能显著提高麻疯树幼苗对盐渍的适应, 主要因为其提高了植株的抗氧化能力、光合作用、渗透调节能力。

关键词: 麻疯树; 多效唑喷施; 幼苗; 耐盐性

Physiological mechanism of foliage spraying paclobutrazol on increasing salt tolerance of *Jatropha curcas* seedlings

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Abstract: Soil salinity is one of the major abiotic stresses with adverse effects on plant productivity and quality. As salt constantly accumulates in soil with irrigation, soil salinity becomes one of the greatest problems in agricultural production through out the world. *Jatropha curcas*, as the first referent tree in development of China biological diesel oil and a hotspot of the research in the world facing global energy crisis recently, has a very high environmental adaptation. However, few studies have been reported on the mechanism of salt adaptation and chemical regulation of the tree. Improvement of salt tolerance to utilize extensive salt soil is an important issue in both agricultural and biological sciences. Moreover, salt tolerance improved by chemical regulation used to be considered as an economic and effective method. Paclobutrazol (PP₃₃₃) causes metabolic and structural changes in plant, and improves plant stress tolerance. Therefore, In order to evaluate the effects of foliage spraying PP₃₃₃ on salt tolerance of *J. curcas* seedlings, the plant dry weight, root/shoot ratio, water content, the leaf cell ultrastructure, chlorophyll content, the photosynthesis, the osmotic adjustment ability and the antioxidation ability of *J. curcas* seedlings treated with 200 mmol/L NaCl for 10 d with foliage spraying in 600 mg/L of PP₃₃₃ in greenhouse were investigated. The results obtained were as follows: (1) 200 mmol/L NaCl stress significantly decreased the seedling dry weight, water content and root/shoot ratio, and drastically increased the leaf electrolyte leakage. However, compared with the plant treated only with NaCl, the plant dry weight, root/shoot ratio and water contents of the seedlings treated with foliage spraying in 600 mg/L of PP₃₃₃ under NaCl stress were all increased significantly, and their leaf

基金项目: 国家支撑项目(2008BAD95B05); 国家公益性行业项目(200903001-05)

收稿日期: 2010-07-07; 修订日期: 2010-10-26

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electrolyte leakage percentage was decreased significantly. (2) The photos of transmission electron microscope indicated that the leaf cells of *J. curcas* contained distinct starch grains, that elliptic chloroplasts clung closely to cell wall under control and that plasmolysis phenomenon and cell membrane damage of leaf cells under 200 mmol/L NaCl were obviously visible. Moreover, NaCl-treated chloroplasts did not contain starch grains but formed osmophilic globules. However, in *J. curcas* seedlings treated with salt and with foliage spraying in 600 mg/L of PP₃₃₃, plasmolysis phenomenon did not occur, osmophilic globule numbers decreased and osmophilic globule volume diminished evidently. (3) *J. curcas* seedlings treated with 200 mmol/L NaCl showed weak osmotic adjustment ability (0.06 MPa) in leaves. However, the seedlings treated with foliage spraying in 600 mg/L of PP₃₃₃ significantly displayed evident promotion in osmotic adjustment ability (0.49 MPa in leaves of non-NaCl treatment and 0.30 MPa in leaves of NaCl treatment). (4) Compared with control, in salt-treated *J. curcas* seedlings, chlorophyll content and net photosynthetic rate (*Pn*) were both decreased evidently. Compared with salt treatment, in the seedlings treated with salt and spraying in 600 mg/L of PP₃₃₃, the chlorophyll content and net photosynthetic rate (*Pn*) were both increased significantly. (5) Compared with control, in salt-treated *J. curcas* seedlings, MDA content and POD activity were greatly enhanced, and SOD and CAT activities were decreased significantly. However, compared with salt treatment, in seedlings treated with salt and foliage spraying in 600 mg/L of PP₃₃₃, MDA content and POD activity were significantly decreased, but SOD and CAT activities increased. To sum up, promotion of salt adaptation of *J. curcas* with foliage spraying in PP₃₃₃ was mainly attributed to enhancement of the photosynthesis, the osmotic adjustment ability and the antioxidation ability.

Key Words: *Jatropha curcas* L. ; foliage spraying in paclobutrazol; seedlings; salt tolerance

土壤盐渍化是影响植物生长、农业生产和生态环境的一个非常重要的非生物胁迫因素^[1-2]。全世界约有9.5亿hm²土地具有不同程度的盐渍化^[3],我国约有3600万hm²各种盐渍土地^[4]。随着我国人口的剧增、城市化以及工业的高速发展,可耕地急剧下降,不合理灌溉、耕作又造成了大量良田的次生盐渍化^[5-6]。如何利用大面积的盐碱地,提高作物耐盐性是农业与生物科学的一个重要研究课题。使用化学诱抗剂,改变植物的生理活动,提高其抗逆性和产量不失为一种简单易行、有效的方法^[7-8]。多效唑(PP₃₃₃)系20世纪80年代新推出的一种高效、低毒的植物生长调节剂,能够抑制植物的纵向生长,而促进其横向生长,同时提高植物的抗逆性^[9-10]。最近的研究结果^[11]表明,600 mg/L PP₃₃₃浸种处理能显著提高麻疯树根、叶的K⁺含量,降低根、叶的盐分,维持体内的离子平衡;同时改善光合作用,显著促进盐胁迫下麻疯树幼苗的生长。相比浸种,在植物不同生长阶段喷施植物生长调节剂往往目的性更强,是更常用、更有效的农艺措施^[10,12-13]。

麻疯树(*Jatropha curcas* L.)在中国种植面积广阔,环境适应性强,生长迅速,生命力强,作为中国发展生物柴油的首选树种,在全球能源危机的今天日益成为公众关注的焦点^[14]。但有关麻疯树对盐渍适应及调节方面的国内外研究并不多^[15-16]。因此,本试验主要探讨了盐胁迫下PP₃₃₃喷施对麻疯树幼苗生长、细胞的超微结构、光合作用、渗透调节能力和抗氧化系统的效应。揭示PP₃₃₃调节麻疯树耐盐性的效应及其机制,为提高麻疯树的耐盐性、麻疯树的盐土种植推广和开发利用提供理论依据。

1 材料和方法

1.1 材料培养与处理

供试麻疯树(*Jatropha curcas* L.)品种为“南油1号”,其种子采自南京农业大学海南滩涂农业研究所。将种子用蒸馏水浸泡24 h,去壳播种于装有蛭石的穴盘中,于光照培养箱中育苗,昼/夜温度为30℃/25℃,光照度1500 lx,相对湿度40%,每天视情况喷水几次,保持基质湿润。培养第6天,挑选生长一致的幼苗,移栽至南京农业大学温室中进行砂培,每处理重复6次,每盆2株。采用1/2 Hoagland培养液浇灌。

缓苗后,傍晚时刻用600 mg/L PP₃₃₃喷施,以蒸馏水喷施为对比。次日傍晚即转入不同处理,试验分为4个处理:(1)对照(CK)为蒸馏水喷施的,继续用不含NaCl的1/2 Hoagland培养液处理;(2)盐处理(SCK)为

蒸馏水喷施的,再转入含 200 mmol/L NaCl 的 1/2 Hoagland 培养液处理;(3) PP₃₃₃ 喷施(P₆₀₀)为 600 mg/L PP₃₃₃ 喷施的,继续用不含 NaCl 的 1/2 Hoagland 培养液处理;(4) PP₃₃₃ 喷施后盐处理(SP₆₀₀)为 600 mg/L PP₃₃₃ 喷施的,再转入含 200 mmol/L NaCl 的 1/2 Hoagland 培养液处理。试验过程中,不同处理的培养液浇灌方式均采用砂面浇灌法,为减少因蒸腾而使得砂培介质中处理液离子浓度变化幅度,处理液浇灌量为砂子持水量的 4 倍,每天浇灌 1 次,约 3/4 的溶液流出,从而将积余的盐冲洗掉,以保持不同处理液成分的恒定。试验处理期间,采用自然光照培养,日温 27—35℃,夜温 24—27℃。盐胁迫处理 10 d 取样测定相关指标。

1.2 测定指标及方法

1.2.1 生长指标的测定 盐胁迫结束后用流水冲洗砂子,轻取苗,用蒸馏水将鲜样洗净吸干,称鲜重;每处理抽样 10 株,然后在 105℃ 杀青 15 min 后,于 75℃ 烘干至恒重,称得干重。按下列公式计算干物质积累速率、含水量和根冠比^[17]:

$$\text{植株干物质积累速率}(\text{mgDW/d}) = (\text{处理后干重}-\text{处理前干重})/\text{处理天数}$$

$$\text{含水量}(\%) = [(\text{处理后叶片鲜重}-\text{处理后叶片干重})/\text{处理后叶片干重}] \times 100$$

$$\text{植株根冠比} = \text{根干重}/\text{地上部干重}$$

1.2.2 叶片质膜透性的测定

选倒 4 叶(功能叶)除去表面沾污物,用蒸馏水冲洗 5 次,用干净纱布轻轻吸干叶片表面水分,然后剪成约 1 cm²的小叶片。将剪下的小叶片混合均匀,快速称取鲜样两份,每份 1 g,放入烧杯。一份在常温下放置 15 min 后加入蒸馏水 50 mL。用真空泵抽气 20 min。另一份加入蒸馏水 50 mL,称重,盖上表皿,置于电炉上煮沸 15 min(煮沸时间依不同植物叶片而定),冷却后再称重并加蒸馏水至原重量,继续浸泡叶片。然后将两杯放置室温下浸提 1 h 左右,用 DDS-11D 型电导仪测定两烧杯电导率,同时测定蒸馏水(空白)的电导率。按下列公式计算叶片电解质相对外渗率,以 ELP(%) 表示^[18],每个处理重复 3 次:

$$ELP(\%) = (\text{常温电导率}-\text{空白电导率})/(\text{煮沸电导率}-\text{空白电导率}) \times 100$$

1.2.3 透射电镜制样与观察

选倒 4 叶(功能叶)除去叶脉,用双面刀切成 1 mm×1.5 mm 左右的小块,迅速投入预冷的 3% 戊二醛中,用注射器反复抽真空,使叶片沉底,置于 4℃ 冰箱中固定 12 h,自然干燥,喷镀金属膜,然后用磷酸缓冲液(pH=7.2)洗涤 3 次,每次 20 min,同时抽气直到切块下沉为止。将洗涤过的材料转移至 1% 铁酸中于 4℃ 固定 4 h 后,用蒸馏水洗涤 3 次,每次 20 min,随后用乙醇脱水,Epon812 包埋,LKB-5 型超薄切片机切片,经醋酸双氧铀染色后,用日本日立 H-7650 透射电镜观察并照相,每处理观察 6 个以上视野,每个处理重复 2 次。

1.2.4 叶绿素含量的测定 选倒 4 叶(功能叶)除去叶脉迅速擦净,准确称取 0.125 g,剪成约 0.5 cm 宽的叶片后置于已装好 25 mL 95% 乙醇的容量瓶中,放入 26℃ 恒温箱中避光保存 24 h,待叶肉组织完全变为白色后,将浸提液倒入 1 cm 光径的比色皿内,在 Lambda 25 紫外-可见分光光度计上分别测定 665、649 和 470 nm 处的吸光度值,根据以下公式计算叶绿素(Chl)含量^[18]:

$$Chl(\text{mg/g 鲜重}) = (18.08A_{649} + 6.63A_{665}) \times V/(1000W)$$

式中,V 为提取液体积(mL),W 为材料重(g)。

1.2.5 净光合速率(P_n)的测定

使用美国 Li-Cor 公司 Li-6400 型便携式光合测定系统,在处理的第 10 天(晴天)9:30—11:33 选倒 4 叶(功能叶)测定叶片的净光合速率(P_n)。测定时光强利用 6400-02LED 红蓝光源控制,光量子通量密度为 1200 $\mu\text{mol}\cdot\text{m}^{-2}\text{s}^{-1}$,大气温度为 21.6—23.5℃,大气 CO₂ 浓度变化范围为 411.59—414.52 $\mu\text{mol/mol}$ 。

1.2.6 叶片饱和渗透势(π_{100})和渗透调节能力(OAA)的测定

π_{100} 用冰点降低法测定。选倒 4 叶(功能叶)立即浸入去离子水中饱和 8 h,取出叶片用去离子水冲洗干净,用吸水纸吸净表面水分,立即放入液氮中 30 min,取出置于医用注射器中,室温下融化 50 min,挤出汁液,参照郑青松等^[18]方法用上海医大仪器厂生产的 8P 型全自动冰点渗透压计测定饱和渗透势。

渗透调节能力计算参照郑青松等^[19]方法

$$\Delta\pi_{100} = \pi_{100}^{\mu} - \pi_{100}^s$$

式中, π_{100}^{μ} 为对照叶片的饱和渗透势, π_{100}^s 为处理叶片的饱和渗透势。

1.2.7 丙二醛(MDA)含量和超氧化物歧化酶(SOD)、过氧化物酶(POD)、过氧化氢酶(CAT)等活性测定

同一处理的4棵植株均选倒4叶(功能叶)除去叶脉,用镊子夹住叶子剪碎混匀,用于测定。MDA含量用硫代巴比妥酸(TBA)方法^[18]测定;SOD活性测定采用氮蓝四唑(NBT)法^[18];POD活性测定采用愈创木酚法^[18],以每分钟内 A_{470} 变化0.01的酶量为1个酶活性单位U;CAT活性参照Aebi法^[18]测定,3mL反应体系中含50mmol/L pH7.0磷酸缓冲液1.9mL,45mmol/L H₂O₂1.0mL,0.1mL酶液,以每分钟内 A_{240} 变化0.01的酶量为1个酶活性单位U。

1.3 数据处理

利用Microsoft Excel软件、SPSS13.0软件进行试验数据的统计,采用Duncan检验进行显著性分析。

2 结果与分析

2.1 盐胁迫下PP₃₃₃喷施对麻疯树幼苗生长的影响

与对照(CK)相比,PP₃₃₃喷施(P₆₀₀)不影响植株的干重和叶片电解质外渗率,但是显著提高植株根冠比和含水量;而盐胁迫(SCK)下麻疯树幼苗干重、根冠比、含水量均显著下降,而叶片电解质外渗率(ELP)显著上升;盐胁迫下PP₃₃₃喷施(SP₆₀₀)相比SCK处理,则可显著提高植株幼苗的干重、根冠比和含水量,降低叶片电解质外渗率,SP₆₀₀处理下麻疯树幼苗干重比SCK处理提高13%,根冠比提高26%,含水量提高18%,而电解质外渗率降低33%(表1)。PP₃₃₃喷施幼苗能显著缓解盐胁迫对麻疯树幼苗生长的伤害。

表1 PP₃₃₃喷施对NaCl胁迫下麻疯树幼苗干重、根冠比、含水量和叶片电解质外渗率的影响

Table 1 Effects of foliage spraying PP₃₃₃ on dry weight, root shoot ratio, water contents and leaf electrolyte leakage of *J. curcas* seedlings under NaCl stress

处理 Treatments	干重/g Dry Weights	根冠比 Root/shoot ratio	含水量/% Water contents	叶片电解质外渗率/% Leaf Electrolyte leakage
CK	2.81±0.10 a	0.145±0.015 b	646.22±12.64 c	20.65±3.62 d
SCK (200 mmol/L NaCl)	2.02±0.05 c	0.095±0.005 c	586.78±19.93 d	57.99±5.42 a
P ₆₀₀ (600 mg/L PP ₃₃₃)	2.99±0.15 a	0.185±0.014 a	786.65±19.75 a	23.26±3.87 c
SP ₆₀₀ (SCK+P ₆₀₀)	2.29±0.08 b	0.120±0.009 bc	692.29±22.34 b	38.58±4.97 b

表中同一列标记相同的小写字母表示它们在5%水平上不存在显著性差异

2.2 PP₃₃₃喷施对NaCl胁迫下麻疯树幼苗叶片细胞超微结构的影响

透射电镜照片(图1)显示,对照处理(CK)的麻疯树叶肉细胞结构正常,无质壁分离现象(a),叶绿体紧贴细胞壁,外膜平滑清晰(b),叶绿体内含有少量淀粉粒(c)(图1A1,A2);盐胁迫(SCK)下,细胞质壁分离现象严重(d),而且细胞膜有断裂的迹象(e),即膜的完整性遭到破坏,叶绿体中几乎无淀粉粒,但有较多体积较小的嗜锇颗粒(f)(图1B1,B2);PP₃₃₃喷施(P₆₀₀)处理的细胞结构与CK无明显差别(图1A1,A2,C1,C2);盐胁迫下PP₃₃₃

喷施(SP₆₀₀)的细胞结构无质壁分离现象(g),嗜锇颗粒数目也大为减少,且体积变小(h)(图1D1,D2),叶绿体紧贴细胞壁(i)(图1D1,D2)。可见,PP₃₃₃喷施显著能减轻盐胁迫对麻疯树幼苗叶片细胞超微结构的破坏。

2.3 PP₃₃₃喷施对NaCl胁迫下麻疯树幼苗叶绿素含量和净光合速率(P_n)的影响

盐胁迫(SCK)下的幼苗叶绿素(Chl)含量和净光合速率(P_n)值比对照(CK)的均显著下降(图2);与对照相比,PP₃₃₃喷施(P₆₀₀)处理显著提高了植株的Chl含量和 P_n ;同样在NaCl胁迫下,PP₃₃₃喷施(SP₆₀₀)的Chl含量和 P_n 显著高于SCK的,但相比CK,是显著降低的。这表明无论是盐胁迫还是非盐胁迫下,PP₃₃₃喷施均能提高幼苗叶片的Chl含量和 P_n ,改善幼苗的光合作用。

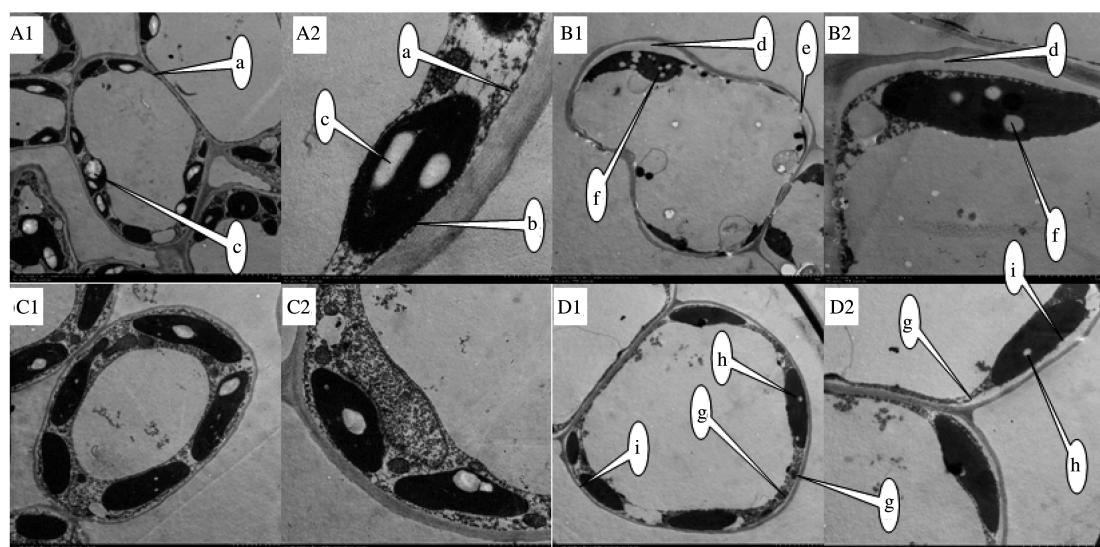


图1 PP₃₃₃喷施对NaCl胁迫下麻疯树幼苗叶片细胞超微结构的影响

Fig. 1 Effects of foliage spraying PP₃₃₃ on the leaf cell ultrastructure of *J. curcas* seedlings under NaCl stress

A1, A2: CK; B1, B2: SCK; C1, C2: P600; D1, D2: SP600

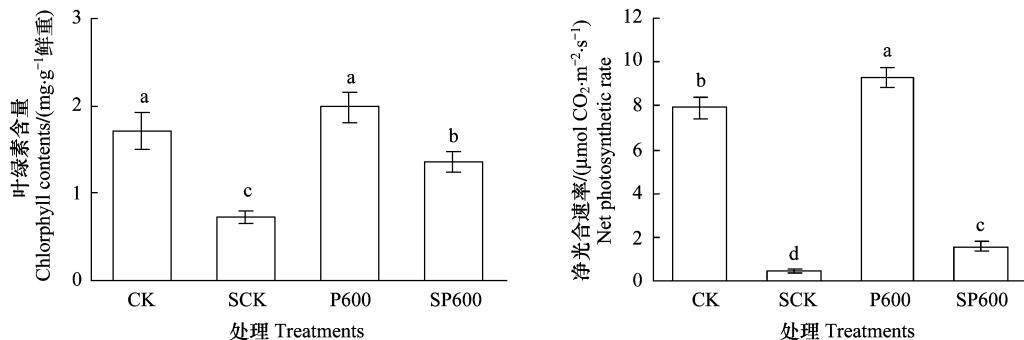


图2 PP₃₃₃喷施对NaCl胁迫下麻疯树幼苗叶片叶绿素含量和净光合速率的影响

Fig. 2 Effects of foliage spraying PP₃₃₃ on leaf chlorophyll content and net photosynthetic rate of *J. curcas* seedlings under NaCl stress

2.4 PP₃₃₃喷施对NaCl胁迫下麻疯树幼苗叶片渗透调节能力的影响

如图3所示,200 mmol/L NaCl胁迫(SCK)10 d,麻疯树幼苗显示出其具有渗透调节能力,但是较弱;而600 mg/L PP₃₃₃喷施(P₆₀₀)处理的幼苗则具有较强的的渗透调节能力,而盐胁迫下PP₃₃₃喷施(SP₆₀₀)处理的渗透调节能力比SCK显著提高,提高了近14倍。可见,PP333喷施显著增强盐胁迫下麻疯树幼苗叶片的渗透调节。

2.5 PP₃₃₃喷施对NaCl胁迫下麻疯树幼苗叶片MDA含量和保护酶(SOD、POD和CAT)活性的影响

如图4所示,相比对照(CK),200 mmol/L NaCl胁迫(SCK)下麻疯树幼苗叶片SOD和CAT活性显著下降,但叶片丙二醛(MDA)含量和POD活性显著增加;200 mmol/L NaCl胁迫下600 mg/L PP₃₃₃喷施(SP₆₀₀)处理的叶片MDA含量相比CK和600 mg/L PP₃₃₃喷施(P₆₀₀)处理要显著增加,但显著低于SCK。SP₆₀₀处理的植株SOD活性虽低于CK,但与P₆₀₀处理的无显著差异,且显著高于SCK的。SP₆₀₀处理的POD活性比SCK的显著下降,比CK的也显著降低,但显著高于P₆₀₀处理的。SP₆₀₀处理的幼苗叶片CAT活性要显著高于SCK的,但低于CK和P₆₀₀处理的。

3 讨论

刺槐种子经250 mg/L PP₃₃₃浸种处理,可提高其叶片的含水量,增强其耐盐性^[20],施用400 mg/L PP₃₃₃多

效唑(溶液外施)后,能显著增加高羊茅生物量,延长根长,促进叶片可溶性糖的积累,保持更高的叶片含水量,减少MDA的生成,从而在一定程度上缓解了高羊茅的盐害^[21]。本研究表明,600 mg/L PP₃₃₃喷施处理明显增强200 mmol/L NaCl胁迫下麻疯树幼苗的光合作用,提高植株干重和含水量,从而提高麻疯树幼苗对盐渍的适应性。

盐胁迫引起的细胞膜透性的变化会导致细胞间和细胞内各种微环境发生改变,从而引起膜和基质间的平衡丧失,各种代谢过程失调,最终导致植物伤害^[22-23]。郑世英等^[24]发现高盐胁迫下玉米的质膜、液泡膜、线粒体和叶绿体等均受到严重的破坏。各种生物膜受到破坏,叶肉细胞发生严重的质壁分离,严重时发生细胞壁断裂,甚至整个细胞溶解。盐胁迫下青钱柳幼苗叶片叶绿体膜消失,类囊体片层结构肿胀,叶绿体降解,叶绿体中淀粉粒水解消失,嗜锇颗粒增大增多。细胞核核膜消失,核染色质出现凝聚。并且细胞还伴随有质壁分离现

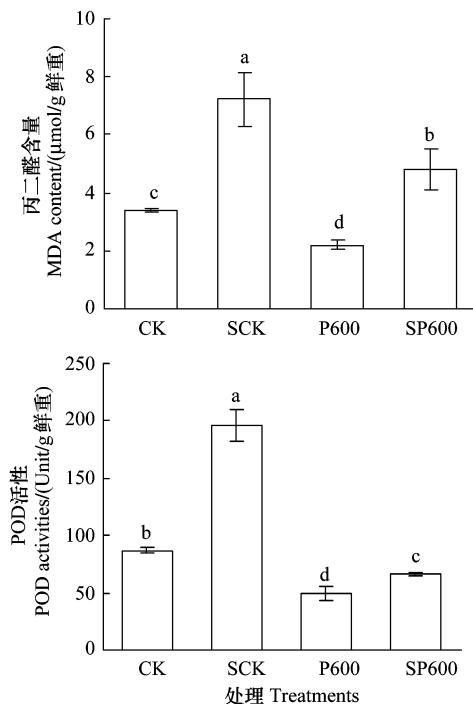


图4 PP₃₃₃喷施对NaCl胁迫下麻疯树幼苗叶片MDA含量以及SOD、POD和CAT活性的影响

Fig. 4 Effects of foliage spraying PP₃₃₃ on malondialdehyde content and SOD, POD and CAT activities of *J. curcas* seedlings leaves under NaCl stress

象的发生^[25]。本研究首次探讨了PP₃₃₃喷施处理对NaCl胁迫下麻疯树幼苗细胞膜透性和叶片细胞超微结构的影响。发现200 mmol/L NaCl处理下,麻疯树幼苗叶片细胞质壁明显分离,细胞膜明显破损,出现较多的嗜锇颗粒;PP₃₃₃喷施明显缓解麻疯树幼苗叶片细胞超微结构的伤害。具体表现为质壁分离现象解除,细胞膜完整性改善,嗜锇颗粒变小,数目也大为减少,从而降低降低了膜透性,叶片电解质外渗率大大降低。

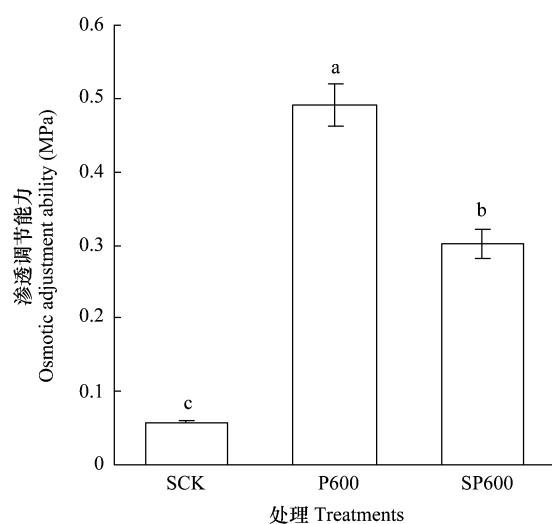


图3 PP₃₃₃喷施对NaCl胁迫下麻疯树幼苗叶片渗透调节能力的影响

Fig. 3 Effects of foliage spraying PP₃₃₃ on osmotic adjustment ability of *J. curcas* seedlings leaves under NaCl stress

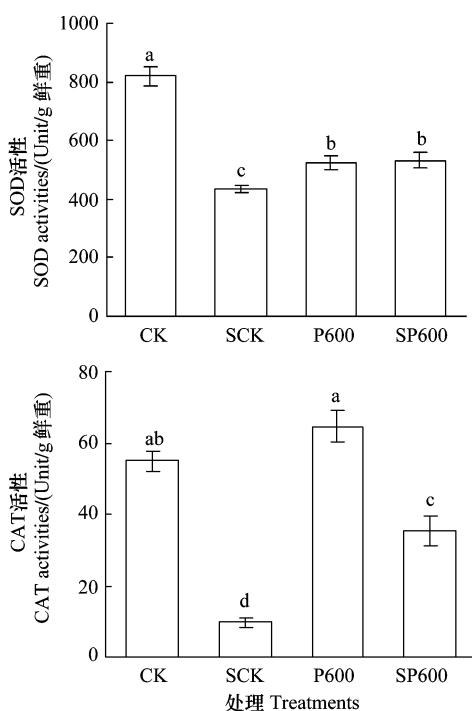


图3 PP₃₃₃喷施对NaCl胁迫下麻疯树幼苗叶片渗透调节能力的影响

Fig. 3 Effects of foliage spraying PP₃₃₃ on osmotic adjustment ability of *J. curcas* seedlings leaves under NaCl stress

PP₃₃₃处理还显著增加盐胁迫和非盐胁迫下长春花体内的过氧化氢酶(CAT)的活性^[10]。Manivannan等^[26]发现PP₃₃₃处理能提高NaCl胁迫下SOD、CAT和POD酶活性。研究结果表明,200 mmol/L NaCl胁迫下,喷施PP₃₃₃显著提高植株叶片SOD和CAT活性。这个结论与前人的结果^[10,27]是一致的。POD酶活性与植物的代谢强度及抗性及植物的衰老都有一定关系。POD的作用具有双重性,一方面,POD可在逆境或衰老初期表达,可清除H₂O₂,表现为保护效应,为细胞活性氧保护酶系统的成员之一;另一方面,POD也可在逆境或衰老后期表达,参与活性氧的生成、叶绿素的降解,并能引发膜脂过氧化作用,表现为伤害效应,是植物体衰老到一定阶段的产物,甚至可作为衰老指标。一般认为其主要作用在于后者^[8,26]。本研究不同处理下POD的活性变化不同于SOD和CAT,200 mmol/L NaCl处理10 d对麻疯树幼苗来说,属于重度胁迫,叶片明显褪绿,POD活性上升125%,提前呈现衰老特征,喷施PP₃₃₃则显著降低植株叶片POD活性。据此认为,PP₃₃₃处理显著提高麻疯树耐盐性很可能也是受到上述抗氧化代谢调节的结果,从而使得植株抗氧化能力提高,大大降低盐胁迫下植株体内的膜脂过氧化,使得植株质膜透性降低,维持膜对离子的较高的选择透过性,从而维持植株叶片光合作用等生理功能。

渗透调节是植物的重要抗逆生理机制^[8,17]。目前已知,许多植物都具有渗透调节能力,不同植物及其品种渗透调节能力的大小不同,渗透调节能力的大小还随环境因素的变化而变化^[27]。而本研究中首次对麻疯树幼苗进行的渗透调节能力研究结果表明,200 mmol/L NaCl这样的重度盐处理下,其叶片渗透调节能力仅仅0.06 MPa。然而,喷施PP₃₃₃则显著提高植株叶片的渗透调节能力,达到0.49 MPa,盐胁迫下,喷施PP₃₃₃的麻疯树幼苗叶片渗透调节能力达到0.30 MPa。渗透调节能力的提高,从而维持细胞膨压,维持叶片的生长速率,提高抗逆性^[8,23]。

总之,从本研究中可以得出结论,对于麻疯树这个首选的生物柴油树来说,喷施600 mg/L PP₃₃₃能够显著缓解其在NaCl胁迫下的细胞伤害程度,保持膜的稳定,促进其光合作用,改善抗氧化代谢,从而提高幼苗干物质积累,是提高其耐盐性的有效方法之一。

致谢:本文中麻疯树叶片渗透调节能力的测定得到南京农业大学生命科学学院於丙军教授和周强博士的帮助,电镜部分试验得到南京农业大学生命科学实验教学中心胡冰老师的帮助。

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*《生态学报》2009 年在核心版的 1964 种科技期刊排序中总被引频次 11764 次, 全国排名第 1; 影响因子 1.812, 全国排名第 14; 第 1—9 届连续 9 年入围中国百种杰出学术期刊; 中国精品科技期刊

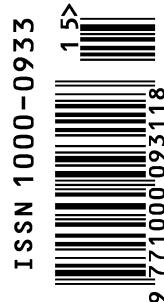
编辑部主任 孔红梅

执行编辑 刘天星 段 靖

生态学报
(SHENGTAI XUEBAO)
(半月刊 1981 年 3 月创刊)
第 31 卷 第 15 期 (2011 年 8 月)

ACTA ECOLOGICA SINICA
(Semimonthly, Started in 1981)
Vol. 31 No. 15 2011

编 辑	《生态学报》编辑部 地址: 北京海淀区双清路 18 号 邮政编码: 100085 电话: (010) 62941099 www. ecologica. cn shengtaixuebao@ rcees. ac. cn	Edited by Editorial board of ACTA ECOLOGICA SINICA Add: 18, Shuangqing Street, Haidian, Beijing 100085, China Tel: (010) 62941099 www. ecologica. cn Shengtaixuebao@ rcees. ac. cn
主 编	冯宗炜	Editor-in-chief FENG Zong-Wei
主 管	中国科学技术协会	Supervised by China Association for Science and Technology
主 办	中国生态学学会 中国科学院生态环境研究中心 地址: 北京海淀区双清路 18 号 邮政编码: 100085	Sponsored by Ecological Society of China Research Center for Eco-environmental Sciences, CAS Add: 18, Shuangqing Street, Haidian, Beijing 100085, China
出 版	科学出版社 地址: 北京东黄城根北街 16 号 邮政编码: 100717	Published by Science Press Add: 16 Donghuangchenggen North Street, Beijing 100717, China
印 刷	北京北林印刷厂	Printed by Beijing Bei Lin Printing House, Beijing 100083, China
发 行	科学出版社 地址: 东黄城根北街 16 号 邮政编码: 100717 电话: (010) 64034563 E-mail: journal@ cspg. net	Distributed by Science Press Add: 16 Donghuangchenggen North Street, Beijing 100717, China Tel: (010) 64034563 E-mail: journal@ cspg. net
订 购	全国各地邮局	Domestic All Local Post Offices in China
国外发行	中国国际图书贸易总公司 地址: 北京 399 信箱 邮政编码: 100044	Foreign China International Book Trading Corporation Add: P. O. Box 399 Beijing 100044, China
广告经营 许 可 证	京海工商广字第 8013 号	



ISSN 1000-0933
CN 11-2031/Q

国内外公开发行

国内邮发代号 82-7

国外发行代号 M670

定价 70.00 元